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Synthesis and antimicrobial activity of 7α -amino-23,24-bisnor- 5α -cholan-22-ol derivatives

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Abstract—A series of 7α -aminobisnorsteroids were synthesized and their in vitro antimicrobial activity was evaluated regarding Gram-positive and Gram-negative bacteria. The stereoselective reductive amination of 7-ketosteroid 3 with NH₄OAc, in the presence of NaBH₃CN, afforded a high yield of 7α -aminosteroid 4. The 3,7-diaminobisnorsteroids were obtained by the reductive amination of 4 with NH₄OT_f, Boc-spermidine, and Boc-spermine. 3α , 7α -Diaminobisnorsterol dihydrochloride 15 showed the highest antimicrobial activity against *Streptococcus pyogenes* 308A with a MIC value of 1.6 μg/mL. Hemolytic activities of the compounds 13–20 were determined. Compound 13 showed MHC value at $100 \mu g/mL$.

Resistance against man-made antibiotics in human pathogens has increased tremendously, thus posing a constant challenge to drug designers. Aminosteroids, compounds that possess an amino group in a steroid scaffold, have recently been recognized as potential antibiotics. The most important example of such an aminosteroid is squalamine (Fig. 1), a polyamine-steroid conjugate isolated from the tissue of Squalus acanthias. Squalamine, which displays antimicrobial activity against Gram-positive and Gram-negative bacteria, possesses antiangiogenic activity and exhibits low hemolytic activity.² Recently, seven new aminosterols related to squalamine were isolated and found to be more potent than squalamine.³ The high potency and low natural abundance of squalamine has prompted efforts toward its synthetic production resulting in a multi-step total synthesis, although in low overall yield.⁴ Analogues of squalamine have been prepared and some of these compounds display stronger activity against bacteria, fungi, and protozoa than squalamine.⁵ Hence, focus has been diverted away from the total synthesis of squalamine to the synthesis of its analogues to eliminate the long,

multi-step, low-yielding syntheses of these potential antibiotics. Toward this end, a variety of aminosteroids have been synthesized for the investigation of their structure–activity relationship (SAR). Recently, a series of cholesterol-based 3-amino and 7-aminosterols were synthesized and found to be active against microbes.⁶ In one study, an amino group was attached to cholesterol through oximation and reduction or azide formation and reduction to synthesize 7α and 7β -spermidinylcholesterol. But these methods required multistep process and overall yields were not so high.⁶ In another study, amphiphilic 3α- and 3β-amino derivatives were synthesized from 5α-cholestan-3β-ol through a multi-step process involving mesylation, bromination, azide formation, and reduction which provided the final products in low overall yields. In contrast to these multi-step methods, employing a reductive amination in the synthesis of aminosteroids offers advantages potentially leading to one-step syntheses of aminosterols in high yields. 8,9 As part of a research program directed toward the development of aminosteroid antibiotics, we have synthesized a series of aminosteroids from 3-keto-23,24-bisnorchola-4-en-22-ol and various amines using a reductive amination approach in order to examine the SAR against bacteria. Our study has focused on the significance of the functional group (hydroxyl, fluorine and amino) at C-7 and the amino or polyamino

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 H_3 NH₉
 H_3 NH₉
 H_4 NH₉

Figure 1. Squalamine and its analogues.

group at C-3, along with the relative stereochemistry at these two positions. Recently, we reported on the synthesis and antibacterial activity of 7-fluoro-3-aminosteroids. ¹⁰ In a continuation of our efforts, we now report on the synthesis and antibacterial activity of 7α -aminobisnorsteroids.

For the synthesis of 7α -aminobisnorsteroids, the requisite starting material, 3-dioxolane-22-tert-butyldimethylsilyloxy-23,24-bisnor-5α-cholan-7-one (3), was prepared from commercially-available 3-keto-23,24-bisnorchola-4-en-22-ol by the previously reported procedure. 11 The stereoselective reductive amination of 3-keto and 7-ketosteroids in order to synthesize 3α , 3β and 7α -isomers, respectively, has been previously reported by us. 8 The versatility of this reductive amination protocol was extended to 23,24-bisnor-5α-cholane 3 to introduce amine functionality at C-7 by reacting compound 3 with NH₄OAc in the presence of NaBH₃CN to give the 7αaminobisnorsteroid. After the groups at C-3 and C-22 were deprotected with pTSA, the free amino group at C-7 was reacted with di-tert-butyl dicarbonate to give 3-keto-7α-(*tert*-butyloxycarbonyl)amino-23,24-bisnor-5αcholan-22-ol 4 in 72% yield. The 3-keto group of 4 was reduced with K-selectride to yield 3α-hydroxy compound 5 in 85% yield, or with NaBH4 which afforded the 3β-hydroxy compound 6 in 78% yield. The stereoselective introduction of an amino group at C-3, with the configurations 3α and 3β was dependent on the selection of a reducing reagent. Smaller reagents like NaBH3CN provided the 3β isomer and bulky reagents like sodium triacetoxyborohydride [NaBH(OAc)₃] and sodium tris[2-(ethylhexanoic)]borohydride [NaBH(OEh)₃]^{8a} afforded 3α isomer predominantly.^{8,12} Thus, amine precursors such as NH₄OT_f, Boc-spermidine, and Bocspermine could be reacted with 4 in the presence of NaBH(OEh)₃ to afford 7 (76%), 9 (93%), and 11 (91%) while the reaction of 4 with the same amine precursors using NaBH₃CN produced 8 (62%), 10 (60%), and 12 (61%) as shown in Scheme 1. The structures of the obtained compounds were characterized by NMR spectroscopy and elemental analysis. 13 The ¹H NMR of 9 showed 3α -NH and 7α -NH protons at δ 4.61 and 4.80, respectively, whereas ¹H NMR of 10 revealed 3β-NH and 7α -NH proton at δ 4.60 and 4.73, respectively.¹³ The chemical shifts observed were similar to those found previously.8b Deprotection of the Boc group in compounds 5-12 with hydrochloric acid in situ generated from the reaction of thionyl chloride with methanol proceed smoothly in dichloromethane to provide the corresponding hydrochloride salts 13–20 quantitatively. Recrystallization of the resulting hydrochloride salts in acetone–methanol afforded pure 13–20. The sulfation of the hydroxyl group at C-22 in 9 and 10 with SO₃–pyridine complex in pyridine at room temperature, followed by treatment with hydrochloric acid yielded 21 and 22 in 94% and 92% yield, respectively.

Ten synthesized 7α -aminobisnorsteroids 13–22 were evaluated along with compounds 1 and 2 for antimicrobial activity against the strains of three Gram-positive and five Gram-negative bacteria as previously described. 10,11 and the MIC (minimum inhibitory concentration) values are summarized in Table 1. The structures of 1 and 18 are similar in all respects including stereochemistry, except for the functional group at C-7. Previously, we examined the effect of substituents by introducing the fluorine atom at C-7 in 2, which showed more potency than the hydroxyl analogue 1.10 Although the 3-amino-7α-hydroxy-23,24-bisnor-5α-cholane was inactive (with a MIC of 100.0 µg/mL) against the tested strains, ¹² the 3-hydroxy-7α-amino-23,24-bisnor-5α-cholanes 13 and 14 showed antimicrobial activity against Streptococcus pyogenes 308A, S. pyogenes 77A, Staphylococcus aureus 503, and Escherichia coli DC2. The facial amphiphilic 3α,7α-diaminobisnorsteroid 15 was the most potent in this series with a MIC as low as 1.6 μg/mL against S. pyogenes 308A. The 3β isomer 16 was less potent than the 3α isomer 15 with a MIC of 6.3 µg/mL against Gram-positive strains. Moreover, it was observed that the 3α-hydroxy isomer 13 was more potent than the 3β-isomer 14, and in the same manner the 3α -amino isomer 15 was more active than the 3β -isomer 16. The introduction of a spermidine moiety at C-3 in a bisnorsteroid did not further enhance activity against any of the tested bacterial strains. Compounds 1, 2, and 18 have a similar structure except for the functional group at C-7, and a comparison of the activity of these compounds shows that an amino substituent only marginally improves activity while a fluorine dramatically enhances potency.¹⁰ The sulfation at C-22 in 17 gave analogue 21 which showed twice the activity as 17 against S. pyogenes 308A, S. pyogenes 77A, S. aureus 503, and *Pseudomonas aeruginosa* 9027. The 3β isomer 22, however, was less active. The spermine analogues 19 and 20 showed enhanced activity against S. pyogenes 308A, S. aureus 503, and P. aeruginosa 9027 compared to the spermidine analogues 17 and 18. The 3β-spermine isomer 20 was more active than the 3α -spermine isomer against S. pyogenes 308A, S. aureus 503, and P. aeruginosa 9027. Compound 15 was moderately active against Salmonella typhimurium and Enterobacter cloacae 1321E as

Scheme 1. Synthesis of 7α -aminosteroids. Reagents: (i) NH₄OAc, NaBH₃CN, THF-MeOH; (ii) pTSA, acetone; (iii) Boc₂O, MeOH; (iv) K-selectride, THF, -60 °C or NaBH₄, EtOH; (v) NH₄OTf, NaBH(OEh)₃, THF; (vi) amine, NaBH₃CN, THF-MeOH, or amine, NaBH(OEh)₃, THF; (vii) SO₃.py, C₅H₅N; (viii) SOCl₂, MeOH, CH₂Cl₂.

Table 1. In vitro antimicrobial activity (MIC: μg/mL) of 7α-aminobisnorsteroids

Strains	1	2	13	14	15	16	17	18	19	20	21	22
S. pyogenes 308A	25.0	6.3	3.1	6.3	1.6	6.3	12.5	12.5	6.3	3.1	6.3	25.0
S. pyogenes 77A	25.0	6.3	6.3	12.5	3.1	6.3	25.0	25.0	25.0	25.0	12.5	50.0
S. aureus 503	12.5	6.3	3.1	12.5	3.1	6.3	12.5	12.5	12.5	6.3	6.3	25.0
E. coli DC 2	25.0	6.3	12.5	25.0	12.5	25.0	>100	50.0	50.0	50.0	25.0	>100
P. aeruginosa 9027	12.5	6.3	>100	>100	50.0	>100	25.0	25.0	12.5	6.3	12.5	>100
P. aeruginosa 1771M	25.0	3.1	>100	>100	25.0	50.0	>100	>100	25.0	50.0	50.0	>100
S. typhimurium	50.0	>100	>100	>100	25.0	50.0	>100	>100	>100	>100	>100	>100
E. cloacae 1321E	50.0	>100	>100	>100	50.0	50.0	>100	>100	>100	>100	>100	>100

shown in Table 1. Minimal hemolytic concentrations (MHC) of **13–20** were determined in triplicate by the literature method. Most of the 7α -aminobisnor derivatives tested had no significant hemolytic activity up to $100 \ \mu g/mL$. Compound **13** showed MHC at $100 \ \mu g/mL$.

In conclusion, we have synthesized a series of 7α -aminobisnorsteroids by reductive amination in high yield. Our results suggest that the nature and stereochemistry of functional groups exerted a major impact on antimicrobial activity. The 3α -hydroxybisnorsteroid 13 and 3α -aminobisnorsteroid 15 were more active than their 3β -hydroxy and 3β -amino counterparts 14 and 16. This seemed to be a general trend except for 3α -sperminylbisnorsteroid 19 which showed lower activity than its 3β analogue. $3\alpha,7\alpha$ -Diaminobisnorsterol dihydrochloride

15 was determined to be the most potent among the tested 7α -aminobisnorsteroids. The results obtained suggest that the stereochemistry and substituent at C-3 and a 7α -amino group are the crucial determinants of activity. Most of the 7α -aminobisnorsteorids exhibited no hemolytic activity. There was no correlation between MICs and MHCs of individual compounds. The compounds are being checked for antiangiogenic activity and a detailed study will be released in future communication.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2008. 03.042.

References and notes

- (a) Foucalt, C.; Brouqui, P. FEMS Immunol. Med. Microbiol. 2007, 49, 173; (b) D'Costa, V. M.; McGrann, K. M.; Hughes, D. W.; Wright, G. D. Science 2006, 311, 374; (c) Wise, R. J. Antimicrob. Chemother. 2006, 57, 1024; (d) Beovic, B. Int. J. Food Microbiol. 2006, 112, 280; (e) Rice, L. B. Am. J. Infect. Control 2006, 34, S11; (f) Tenover, F. C. Am. J. Med. 2006, 119, S3; (g) Kollef, M. H.; Micek, S. T. Curr. Opin. Infect. Dis. 2006, 19, 161; (h) Richter, S. S.; Heilmann, K. P.; Beekmann, S. E.; Miller, N. J.; Rice, C. L.; Doern, G. V. Clin. Infect. Dis. 2005, 40, 225; (i) Walsh, C. Nature 2000, 406, 775.
- (a) Moore, K. S.; Wehrli, S.; Roder, H.; Rogers, M.; Forrest, J. N., Jr.; McCrimmon, D.; Zasloff, M. Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 1354; (b) Herbst, R. S.; Hammond, L. A.; Carbone, D. P.; Tran, H. T.; Holroyd, K. J.; Desai, A.; Williams, J. I.; Bekele, B. N.; Hait, H.; Allgood, V.; Solomon, S.; Schiller, J. H. Clin. Cancer Res. 2003, 9, 4108; (c) Hao, D.; Hammond, L. A.; Eckhardt, S. G.; Patnaik, A.; Takimoto, C. H.; Schwartz, G. H.; Goetz, A. D.; Tolcher, A. W.; McCreery, H. A.; Mamun, K.; Williams, J. I.; Holroyd, K. J.; Rowinsky, E. K. Clin. Cancer Res. 2003, 9, 2465.
- 3. Rao, M. N.; Shinnar, A. E.; Noecker, L. A.; Chao, T. L.; Feibush, B.; Snyder, B.; Sharkansky, I.; Sarkahian, A.; Zhang, X.; Jones, S. R.; Kinney, W. A.; Zasloff, M. *J. Nat. Prod.* **2000**, *63*, 631.
- (a) Okumura, K.; Nakamura, Y.; Takeuchi, S.; Kato, I.; Fujimoto, Y.; Ikekawa, N. *Chem. Pharm. Bull.* 2003, 51, 1177; (b) Zhang, D.-H.; Cai, F.; Zhou, X.-D.; Zhou, W.-S. *Org. Lett.* 2003, 5, 3257; (c) Kinney, W. A.; Zhang, X.; Williams, J. I.; Johnston, S.; Michalak, R. S.; Deshpande, M.; Dostal, L.; Rosazza, J. P. N. *Org. Lett.* 2000, 2, 2921; (d) Weis, A. L.; Bakes, T.; Alferiev, I.; Zhang, X.; Shao, B.; Kinney, W. A. *Tetrahedron Lett.* 1999, 40, 4863; (e) Jones, S. R.; Selinsky, B. S.; Rao, M. N.; Zhang, X.; Kinney, W. A.; Tham, F. S. *J. Org. Chem.* 1998, 63, 3786;

- (f) Pechulis, A. D.; Bellevue, F. H.; Cioffi, C. L.; Trapp, S. G.; Fojtik, J. P.; McKitty, A. A.; Kinney, W. A.; Frye, L. L. J. Org. Chem. 1995, 60, 5121; (g) Moriarty, R. M.; Enaehe, L. A.; Kinney, W. A.; Allen, C. S.; Canary, J. W.; Tuladhar, S. M.; Guo, L. Tetrahedron Lett. 1995, 36, 5139.
- (a) Chen, W. H.; Shao, X. B.; Moellering, R.; Wennersten, C.; Regen, S. L. Bioconjugate Chem. 2006, 17, 1582; (b) Zhang, D. H.; Cai, F.; Zhou, X. D.; Zhou, W. S. Chin. J. Chem. 2005, 23, 176; (c) Shu, Y.; Jones, S. R.; Kinney, W. A.; Selinsky, B. S. Steroids 2002, 67, 291; (d) Kikuchi, K.; Bernard, E. M.; Sadownik, A.; Regen, S. L.; Armstrong, D. Antimicrob. Agents Chemother. 1997, 41, 1433; (e) Jones, S. R.; Kinney, W. A.; Zhang, X.; Jones, L. M.; Selinsky, B. S. Steroids 1996, 61, 565; (f) Sadownik, A.; Deng, G.; Janout, V.; Regen, S. L.; Bernard, E. M.; Kikuchi, K.; Armstrong, D. J. Am. Chem. Soc. 1995, 117, 6138.
- (a) Choucair, B.; Dherbomez, M.; Roussakis, C.; El kihel, L. Tetrahedron 2004, 60, 11477; (b) Choucair, B.; Dherbomez, M.; Roussakis, C.; El kihel, L. Bioorg. Med. Chem. Lett. 2004, 14, 4213; (c) El Kihel, L.; Choucair, B.; Dherbomez, M.; Letourneux, Y. Eur. J. Org. Chem. 2002, 4075; (d) Fouace, S.; El Kihel, L.; Dherbomez, M.; Letourneux, Y. Bioorg. Med. Chem. Lett. 2001, 11, 3011.
- 7. Sugandhi, E. W.; Slebodnick, C.; Falkinham, J. O., III; Gandour, R. D. Steroids 2007, 72, 615.
- 8. (a) Khan, S. N.; Cho, N. J.; Kim, H.-S. In *Catalysts for Fine Chemical Synthesis*; Robert, S. M., Whittall, J., Eds.; Regio- and Stereo-Controlled Oxidations and Reductions; John Wiley & Sons: Chichester, 2007; Vol. 5, p 175; (b) Khan, S. N.; Cho, N. J.; Kim, H.-S. *Tetrahedron Lett.* 2007, 48, 5189; (c) Khan, S. N.; Bae, S. Y.; Kim, H.-S. *Tetrahedron Lett.* 2005, 46, 7675.
- (a) Loncle, C.; Salmi, C.; Letourneux, Y.; Brunel, J. M. Tetrahedron 2007, 63, 12968; (b) Salmi, C.; Loncle, C.; Vidal, N.; Letourneux, Y.; Brunel, J. M. Eur. J. Med. Chem. 2008, 43, 540; (c) Salmi, C.; Letourneux, Y.; Brunel, J. M. Lett. Org. Chem. 2006, 3, 384; (d) Salmi, C.; Letourneux, Y.; Brunel, J. M. Lett. Org. Chem. 2006, 3, 396.
- Khan, S. N.; Kim, B. J.; Kim, H.-S. Bioorg. Med. Chem. Lett. 2007, 17, 5139.
- (a) Kim, H.-S.; Kwon, K.-C.; Kim, K. S.; Lee, C. H. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 3065; (b) Kim, H.-S.; Choi, B. S.; Kwon, K. C.; Lee, S. O.; Kwak, H. J.; Lee, C. H. *Bioorg. Med. Chem.* **2000**, *8*, 2059.
- Khan, S. N. Ph.D. Thesis, Kyungpook National University, 2007.
- 13. Refer to supplementary information.